

# Comparison of seasonal accumulation by mountain stream algae *Hydrurus foetidus* and *Oscillatoria* sp.

J. TUCHYŇA

*Institute of High Mountain Biology, Žilina University,  
Tatranská Javorina 7, SK-059 56, Slovak Republic;  
e-mail: tuchyna168@gmail.com*

**Abstract.** In the process of biomonitoring, it is crucial to be aware of how species behave and how their metabolic needs influence element accumulation. In this study, we monitored two mountain stream species, *Hydrurus foetidus* and *Oscillatoria* sp. to determine their individual seasonal bio-accumulative abilities and their individual seasonal metabolic traits. Research was carried out at three sites in the foothills of the High Tatras between 2020 and 2023. Samples were collected at monthly intervals and subsequently processed with a fluorescence spectrophotometer. Seasonal changes were largely influenced by the metabolic needs of algae and cyanobacteria. For *Oscillatoria* sp., an increase in S and K was observed during summer, when they reach their metabolic maximum. The metabolic maximum of *H. foetidus* was determined to occur in autumn, based on increased concentration of Cl and Cr. Concentration of Ca and Sr present in *Oscillatoria* sp. exhibited a seasonally dependent pattern of accumulation of these elements during the colder months. The lowest concentrations were found during summer.

**Key words:** algae, cyanobacteria, accumulation of elements, mountain stream

## Introduction

The value of algae as bio-monitors for fresh water has been recognized since the mid-19<sup>th</sup> century (Cohn 1853). Algae are most known as an indicator of organic pollution such as eutrophication (Palmer 1969), however many studies confirm that they can be used also as bio-accumulators of heavy metal pollution and act as their bio-indicator (Zeraatkar *et al.* 2016; Salama *et al.* 2019; Lin *et al.* 2020). The potential of algae for coping with various environmental crises makes them ideal organisms for bio-monitoring water pollution, as well as species with the capacity for removal of pollutants in the process of bioremediation (Roy *et al.* 2022).

Algae have an increased ability to bind elements (Mehta and Gaur 2001). The algae species studied in this research differ in their morphology. *Hydrurus foetidus* of the class Chrysophyceae (golden algae), is a large (up to 10–30 cm), branched, freshwater algae (Wehr and Sheath 2003). Alginates are the main group involved in heavy metal accumulation in brown algae (Kuyucak and Volesky 1989). The negative charge of carboxylic groups plays an important role in binding cations. Polyguluronic and guluronic acids present in alginates (Haug and Smidsrød 1967) showed a high specificity for divalent metal ions such as Pb<sup>2+</sup>, Cu<sup>2+</sup>, Cd<sup>2+</sup>, Zn<sup>2+</sup>, Ca<sup>2+</sup>, ... (Haug *et al.* 1961; Puranik *et al.* 1999). Seki and Suzuki (1998) showed that the biosorption of bivalent metal ions (Fe<sup>2+</sup>, Ca<sup>2+</sup>, S<sup>2+</sup>) by brown algae was due to the presence of carboxylic acids on the alginate groups.

Sheaths of cyanobacteria consist of polysaccharides. Its anionic nature is caused by the presence of uronic acids and/or other charged groups (Mehta and Gaur 2001). Their cell walls provide mainly carboxylic groups for metal binding (Mehta and Gaur 2001; Chojnacka *et al.* 2005). Carboxyl groups serve as a dominant active site for metal ion accumulation. Lipopolysaccharides, lipids, and peptidoglycans are associated with phosphoryl groups. Amino groups are associated with membrane proteins and the peptide component of peptidoglycan (Chojnacka *et al.* 2005).

This research is focussed on differences in accumulation between golden algae *Hydrurus foetidus* and cyanobacteria of the genus *Oscillatoriales* in the mountain stream Javorinka. Selected species for monitoring differ by morphology, physiology, and their life cycle. As such, we predict that changes in element accumulation will be seasonally affected. The goal of the study is to determine how the two species differ in terms of element accumulation and their metabolic needs.

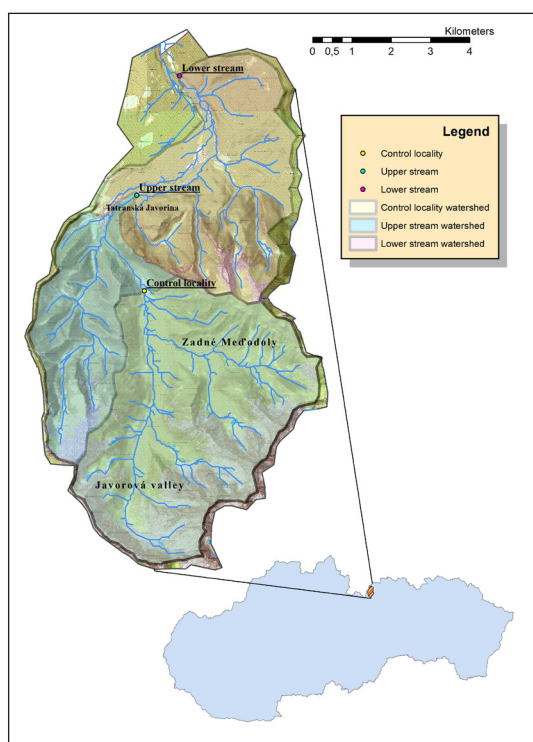
## Material and Methods

### *Location of the research*

Research to monitor the accumulation of elements in algae (*Hydrurus foetidus*, *Oscillatoriales* sp.) is a subsection of broader research undertaken by the Institute of High Mountain Biology of the University of Žilina (IHMB), focused on long-term changes in the levels of elements in the ecosystem of the mountain stream Javorinka.

The Javorinka stream springs at the Žabie Javorové tarn (Javorová dolina, High Tatras) in the northern part of the Eastern Tatras. It is a right-side tributary of the river Biaľka (Poland). The length of its flow is 19.3 km, and the watershed area is 66 km<sup>2</sup> (Bohuš 1996). Javorinka is a high-mountain stream with a snow-rainfall runoff regime. Most of the riverbed consists of clay, or clay-gravel sediments with boulder-like forms. Parent rocks in the Javorinka watershed area are primarily biotic granodiorites, dolomites, limestones, slates, and sandstones.

Research was conducted at three sites along the Javorinka stream, ranging from mountain sites to sub-mountain sites in local villages. The upper-most site was the Control Locality. This sampling site is located at the mouth of the valley Javorová dolina and the valley Zadné Medodoly (N 49.250419° ; E 20.155494° ; 1180 m a.s.l.). It is 1.5 km from the village of Tatranská Javorina (against the flow) and 2.5 km from the second location – the Upper stream. This sampling site is located between the villages of Tatranská Javorina and Podspády (N 49.271164° ; E 20.152952° ; 960 m a.s.l.). The third sampling site - the Lower Stream in the locality of Vojtasová (N 49.292550° ; E 20.169038° ; 850 m a.s.l.). The sampling localities are depicted in Fig. 1.



**Fig. 1.** Map of study area with 3 localities of collecting samples (Lower stream, Upper stream, Control locality). On the map is also shown watersheds corresponding to the exact localities.

#### Field sampling and sample preparation

Sampling was carried out at regular monthly intervals at all three sampling points from January 2022 to December 2023. Samples were collected within a maximum radius of 100 m downstream of the sampling site.

*H. foetidus* was collected by hand due to its macroscopic branched thallus. *Oscillatoriales* were scraped of the rocks using tweezers. Approximately 15 mg of each sample was collected. Samples were stored in plastic bottles with a small amount of water from the stream. In the laboratory, the water-deprived samples were placed on a petri dish and stripped of macroscopic contaminants (invertebrates, larger soil particles). The samples were allowed to dry at room temperature for at least 5 days.

#### Laboratory analyses

The dried samples were homogenized into a fine dust using a CryoMill (Retsch, Germany). It mills each sample for 40 seconds.

Analysis of metallic elements in algae was performed using X-ray fluorescence spectrometry. The samples were analyzed using an ED-XRF Spectrometer DELTA CLASSIC (Innov-X Systems, USA). Each sample was measured for 80 seconds in triplicate; the final value was the average of these measurements. The XRF Spectrometer was calibrated using the INCT-PVTL-6 Polish Virginia Tobacco Leaver standard. Measured were these elements: P, S, Cl, K, Ca, Ti, Cr, Mn, Fe, Co, Ni, Cu, Zn, As, Se, Rb, Sr, Zr, Mo, Ag, Cd, Sn, Sb, Ba, Hg, Pb.

#### Statistical analysis

Only elements that were regularly measured above the detection limit were included in the statistical analysis. Statistica 8 (Data Analysis Software System; Statsoft, Inc.) was used for statistical analysis of the data. Normality of the data was tested by the Shapiro-Wilk test. Comparisons of individual items were made with the nonparametric Kruskal-Wallis H test (K-W H) because the variances between groups were highly heterogeneous. Values with  $p < 0.05$  were considered statistically significant.

## Results

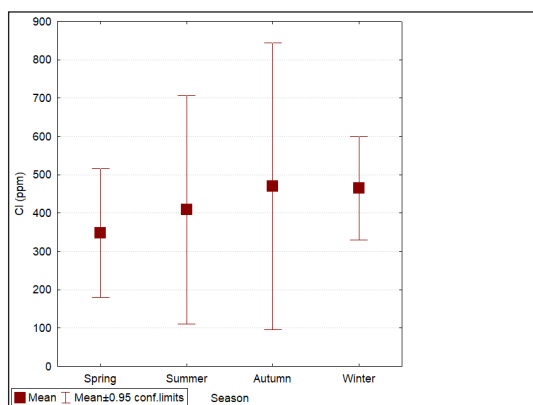
While 26 metallic elements were spectrometrically monitored in the analysed algae and cyanobacteria samples, only 13 elements (S, Cl, K, Ca, Ti, Cr, Mn, Fe, Zn, Rb, Sr, Ba, Pb) were evaluated for statistical analyses (analysis of variance) comparing seasonal accumulation. These are elements regularly measured above the detection limit of the instrument during analysis. The values of other elements were measured irregularly.

In this analysis was observed seasonal variations of the two species individually to better understand metabolic needs of the study organisms. In *H. foetidus*, seasonal differences in Cl and Cr accumulation were significant ( $p < 0.05$ ) (Table 1). Both elements reached their maximum in autumn during the highest biological activity of algae (Figs. 2-3). In *Oscillatoriales*, significant seasonal differences were found for 7 (S, K, Ca, Cr, Mn, Sr, Pb) of the 13 elements tested (Table 2). Both S and K showed maximum concentration in summer, during peak productivity of this genus (Figs. 4-5). Ca and Sr had the lowest mean concentration during summer and the highest during winter. However,

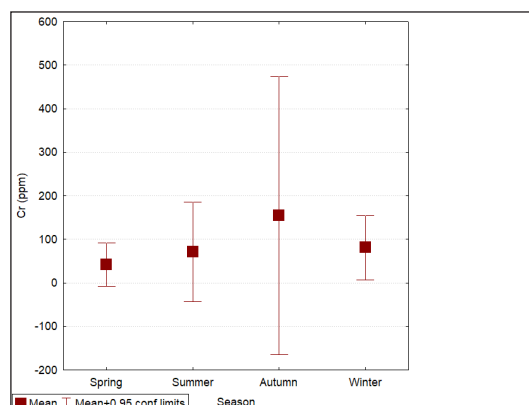
**Table 1.** Characteristic values of analyzed elements accumulated by *Hydrurus foetidus*. Values of elements with significant differences in the spring — winter seasons are highlighted in bold.

	Season	Mean $\pm$ SD (n)	Minimum	Maximum	KW - H	p
S	Spring	4828.56 $\pm$ 1711.09 (18)	2783	9663	3.27	0.3508
	Summer	5575.65 $\pm$ 2510.88 (17)	3159	12584		
	Autumn	5313.71 $\pm$ 1646.52 (14)	1205	8021		
	Winter	5451.67 $\pm$ 1360.9 (9)	4050	8003		
Cl	<b>Spring</b>	<b>344.28 <math>\pm</math> 98.65 (18)</b>	<b>150</b>	<b>519</b>	<b>11.63</b>	<b>0.0087</b>
	<b>Summer</b>	<b>408.29 <math>\pm</math> 152.12 (17)</b>	<b>182</b>	<b>874</b>		
	<b>Autumn</b>	<b>431.5 <math>\pm</math> 225.65 (14)</b>	<b>150</b>	<b>1079</b>		
	<b>Winter</b>	<b>445.89 <math>\pm</math> 129.49 (9)</b>	<b>150</b>	<b>579</b>		
K	Spring	5075.78 $\pm$ 2198.2 (18)	1859	8364	1.15	0.7631
	Summer	5460.76 $\pm$ 2593.61 (17)	1528	9004		
	Autumn	6529.02 $\pm$ 3431.88 (14)	2842	14412		
	Winter	6031.56 $\pm$ 2282.03 (9)	3798	10733		
Ca	Spring	21919.39 $\pm$ 19136.4 (18)	3116	64629	0.74	0.8631
	Summer	23072.71 $\pm$ 28684.6 (17)	3627	107320		
	Autumn	23603.01 $\pm$ 21964.5 (14)	3200	73372		
	Winter	33519.11 $\pm$ 45429.2 (9)	5841	150254		
Ti	Spring	557.56 $\pm$ 744.7 (18)	15	2325	2.79	0.4235
	Summer	640.71 $\pm$ 681.41 (17)	15	1993		
	Autumn	922.14 $\pm$ 1033.41 (14)	15	3346		
	Winter	578.44 $\pm$ 731.01 (9)	48	2207		
Cr	<b>Spring</b>	<b>40.39 <math>\pm</math> 24.98 (18)</b>	<b>13</b>	<b>95</b>	<b>17.75</b>	<b>0.0005</b>
	<b>Summer</b>	<b>71.41 <math>\pm</math> 58.26 (17)</b>	<b>14</b>	<b>187</b>		
	<b>Autumn</b>	<b>157.14 <math>\pm</math> 128.58 (14)</b>	<b>49</b>	<b>445</b>		
	<b>Winter</b>	<b>82.89 <math>\pm</math> 37.96 (9)</b>	<b>41</b>	<b>167</b>		
Mn	Spring	114.16 $\pm$ 78.97 (18)	34.9	277	4.59	0.2037
	Summer	139.65 $\pm$ 85.71 (17)	48	369		
	Autumn	183.57 $\pm$ 95.56 (14)	69	344		
	Winter	149.89 $\pm$ 112.37 (9)	60	362		
Fe	Spring	5335.61 $\pm$ 5717.2 (18)	342	16499	1.89	0.5954
	Summer	6274.29 $\pm$ 5524.72 (17)	802	16099		
	Autumn	8919.93 $\pm$ 8685.21 (14)	820	31326		
	Winter	5702.67 $\pm$ 5610.1 (9)	1164	15331		
Zn	Spring	91.78 $\pm$ 39.99 (18)	26	157	2.75	0.4311
	Summer	99.29 $\pm$ 52.21 (17)	23	181		
	Autumn	106.57 $\pm$ 56.11 (14)	31	188		
	Winter	108.78 $\pm$ 63.76 (9)	23	226		
Rb	Spring	12.37 $\pm$ 8.11 (18)	5.4	29.8	2.51	0.4722
	Summer	13.01 $\pm$ 6.63 (17)	5.8	26.3		
	Autumn	16.86 $\pm$ 11.7 (14)	6.7	43.2		
	Winter	13.13 $\pm$ 7.92 (9)	7.4	30.7		
Sr	Spring	51.85 $\pm$ 44.74 (18)	10	140	0.81	0.8452
	Summer	50.61 $\pm$ 45.03 (17)	11.2	146		
	Autumn	65.32 $\pm$ 58.28 (14)	11.9	188		
	Winter	68.37 $\pm$ 70.43 (9)	13.7	222		
Ba	Spring	159.44 $\pm$ 129.08 (18)	45	443	3.31	0.3456
	Summer	188.24 $\pm$ 117.81 (17)	57	423		
	Autumn	274.5 $\pm$ 213.17 (14)	66	822		
	Winter	190.78 $\pm$ 122.39 (9)	86	447		

	Spring	15.94 ± 4.11 (18)	11	25		
Pb	Summer	17.29 ± 3.95 (17)	12	24	2.38	0.4957
	Autumn	19.43 ± 8.73 (14)	12	46		
	Winter	16.22 ± 3.11 (9)	11	22		



**Fig. 2.** Mean of case coordinates ( $\pm 95$  c. limits) of chlorine concentrations accumulated by *Hydrurus foetidus* during seasons.

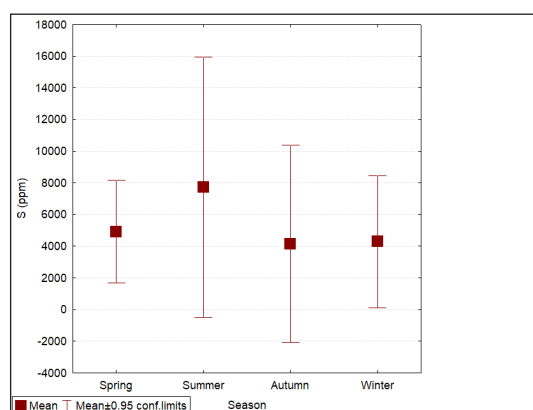


**Fig. 3.** Mean of case coordinates ( $\pm 95$  c. limits) of chromium concentrations accumulated by *Hydrurus foetidus* during seasons.

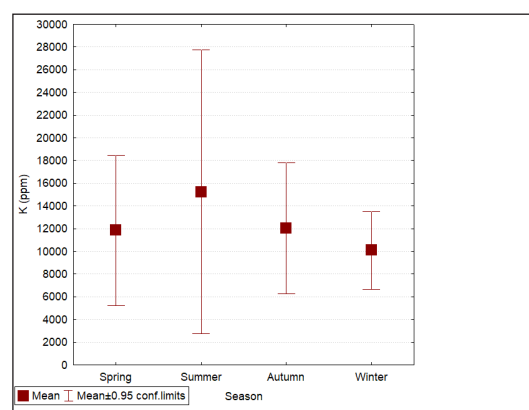
**Table 2.** Characteristic values of analyzed elements accumulated by *Oscillatoria* sp. Values of elements with significant differences in the spring — winter seasons are highlighted in bold.

	Season	Mean ± SD (n)	Minimum	Maximum	KW - H	p
S	<b>Spring</b>	<b>4921 ± 1648.56 (15)</b>	<b>2810</b>	<b>8373</b>	<b>14.51</b>	<b>0.0023</b>
	<b>Summer</b>	<b>7733 ± 4193.48 (18)</b>	<b>2554</b>	<b>18812</b>		
	<b>Autumn</b>	<b>4832 ± 3430.03 (13)</b>	<b>1133</b>	<b>12420</b>		
	<b>Winter</b>	<b>4872 ± 2148.35 (9)</b>	<b>2083</b>	<b>7854</b>		
Cl	Spring	343 ± 77.89 (14)	226	498	6.08	0.1166
	Summer	795 ± 1180.75 (17)	241	5297		
	Autumn	432 ± 209.83 (13)	175	951		
	Winter	392 ± 130.49 (9)	222	620		
K	<b>Spring</b>	<b>11852 ± 3373.94 (15)</b>	<b>7319</b>	<b>18314</b>	<b>11.45</b>	<b>0.0095</b>
	<b>Summer</b>	<b>15250 ± 6366.01 (18)</b>	<b>9077</b>	<b>31970</b>		
	<b>Autumn</b>	<b>14077 ± 6471.48 (13)</b>	<b>9168</b>	<b>33415</b>		
	<b>Winter</b>	<b>9742 ± 1268.43 (9)</b>	<b>8211</b>	<b>12192</b>		
Ca	<b>Spring</b>	<b>72771 ± 61373.1 (15)</b>	<b>10799</b>	<b>169812</b>	<b>10.06</b>	<b>0.0181</b>
	<b>Summer</b>	<b>28633 ± 20976.5 (18)</b>	<b>8215</b>	<b>82027</b>		
	<b>Autumn</b>	<b>39440 ± 21316 (13)</b>	<b>11246</b>	<b>71409</b>		
	<b>Winter</b>	<b>68408 ± 20141.5 (9)</b>	<b>28685</b>	<b>90898</b>		
Ti	Spring	1201.8 ± 966.77 (15)	238	3269	4.25	0.2348
	Summer	1040.39 ± 487.29 (18)	395	2165		
	Autumn	1797.77 ± 1633.31 (13)	593	6986		
	Winter	1003.11 ± 407.65 (9)	465	1566		
Cr	<b>Spring</b>	<b>82.33 ± 84.37 (15)</b>	<b>9</b>	<b>272</b>	<b>9.91</b>	<b>0.0193</b>
	<b>Summer</b>	<b>112.39 ± 94.56 (18)</b>	<b>18</b>	<b>337</b>		
	<b>Autumn</b>	<b>249.08 ± 271.7 (13)</b>	<b>81</b>	<b>1050</b>		
	<b>Winter</b>	<b>92.56 ± 34.22 (9)</b>	<b>61</b>	<b>166</b>		
Mn	<b>Spring</b>	<b>177.27 ± 115.7 (15)</b>	<b>52</b>	<b>406</b>	<b>8.12</b>	<b>0.0468</b>
	<b>Summer</b>	<b>191.28 ± 56.57 (18)</b>	<b>103</b>	<b>264</b>		
	<b>Autumn</b>	<b>252.38 ± 76.73 (13)</b>	<b>156</b>	<b>473</b>		
	<b>Winter</b>	<b>172.78 ± 49.37 (9)</b>	<b>105</b>	<b>264</b>		

Fe	Spring	12133.27 ± 10412.3 (15)	2219	33603	3.53	0.3164
	Summer	10467.72 ± 4685.39 (18)	4235	21454		
	Autumn	14384.85 ± 8257.67 (13)	7104	39947		
	Winter	8999.56 ± 3041.8 (9)	5107	15226		
Zn	Spring	45.53 ± 21.58 (15)	19	81	3.94	0.2677
	Summer	53 ± 14.38 (18)	31	82		
	Autumn	48.38 ± 15.18 (13)	31	84		
	Winter	41.56 ± 12.63 (9)	30	67		
Rb	Spring	20.49 ± 11.91 (15)	8.4	44.1	6.11	0.1059
	Summer	18.21 ± 6.09 (18)	11.8	35		
	Autumn	23.73 ± 7.16 (13)	13.2	32.7		
	Winter	18.54 ± 4.77 (9)	12.7	24.7		
Sr	<b>Spring</b>	<b>116.82 ± 68.58 (15)</b>	<b>23.7</b>	<b>230</b>	<b>9.25</b>	<b>0.0261</b>
	<b>Summer</b>	<b>69.42 ± 59.87 (18)</b>	<b>17.1</b>	<b>190</b>		
	<b>Autumn</b>	<b>115.28 ± 75.67 (13)</b>	<b>20</b>	<b>245</b>		
	<b>Winter</b>	<b>136.56 ± 47.44 (9)</b>	<b>54</b>	<b>194</b>		
Ba	Spring	302.33 ± 184.77 (15)	100	668	5.11	0.1639
	Summer	289.11 ± 103.3 (18)	139	455		
	Autumn	455.08 ± 338.45 (13)	210	1549		
	Winter	260.56 ± 70 (9)	146	380		
Pb	<b>Spring</b>	<b>18.8 ± 7.2 (15)</b>	<b>9</b>	<b>33</b>	<b>9.33</b>	<b>0.0252</b>
	<b>Summer</b>	<b>20.83 ± 4.62 (18)</b>	<b>14</b>	<b>33</b>		
	<b>Autumn</b>	<b>23.85 ± 3.11 (13)</b>	<b>18</b>	<b>31</b>		
	<b>Winter</b>	<b>19 ± 3.87 (9)</b>	<b>13</b>	<b>26</b>		



**Fig. 4.** Mean of case coordinates ( $\pm 95$  c. limits) of sulfur concentrations accumulated by *Oscillatoria* sp. during seasons.



**Fig. 5.** Mean of case coordinates ( $\pm 95$  c. limits) of potassium concentrations accumulated by *Oscillatoria* sp. during seasons.

total maximum was reached in autumn (Figs. 6-7). Mn and Pb had the greatest variance during spring snowmelt season. Maximum concentration of Mn and Pb was reached during autumn (Figs. 8-9). Cr accumulated in great variance from spring until autumn, and similarly to *H. foetidus* it reached its peak during autumn (Fig. 10).

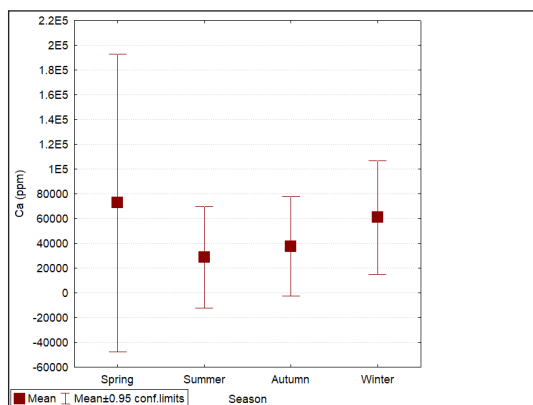
## Discussion

### Chlorine in *Hydrurus foetidus*

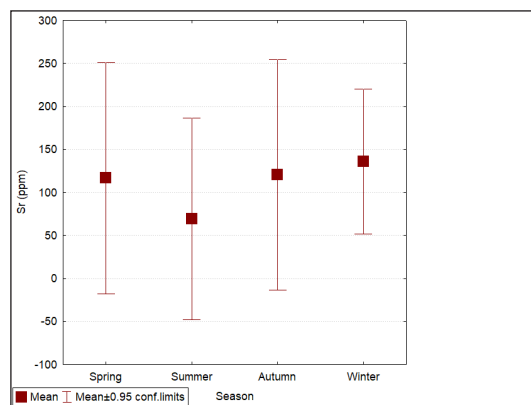
It was observed that many organisms including algae (Gribble 1994; Nzengung *et al.* 1999; La Barre

*et al.* 2010) produce chlorinated compounds as secondary metabolites. Nzengung *et al.* (1999) and Küpper *et al.* (2008) proposed that the transformation of halogenated compounds is processed through sequestration, phytoexudation, partial exudation of the metabolites, phytooxidation of the sequestered phytoexudation products, and assimilation.

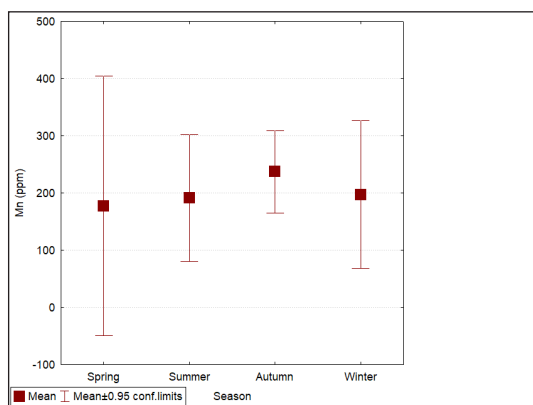
*H. foetidus* experiences its highest biological activity from autumn until late winter (Kann 1978; Klaveness 2019). During this time period, the highest mean Cl concentrations (431.5 ppm in autumn and 445.81 ppm in winter) were also observed. During this period, the highest iodine content as a product of halogenation in the brown algae *Laminaria digitata* was also observed. It can be assumed that



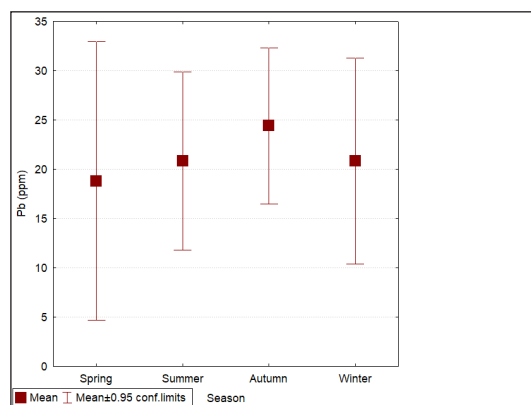
**Fig. 6.** Mean of case coordinates ( $\pm 95$  c. limits) of calcium concentrations accumulated by *Oscillatoria* sp. during seasons.



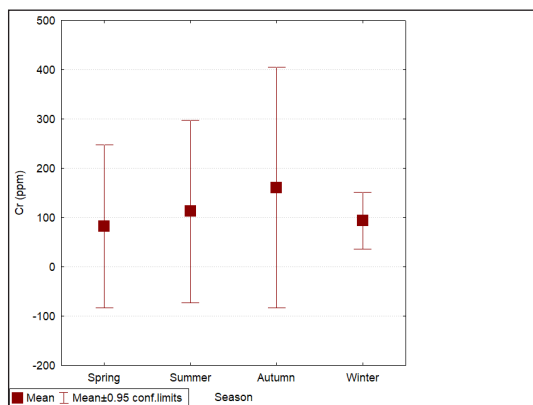
**Fig. 7.** Mean of case coordinates ( $\pm 95$  c. limits) of strontium concentrations accumulated by *Oscillatoria* sp. during seasons.



**Fig. 8.** Mean of case coordinates ( $\pm 95$  c. limits) of manganese concentrations accumulated by *Oscillatoria* sp. during seasons.



**Fig. 9.** Mean of case coordinates ( $\pm 95$  c. limits) of lead concentrations accumulated by *Oscillatoria* sp. during seasons.



**Fig. 10.** Mean of case coordinates ( $\pm 95$  c. limits) of chromium concentrations accumulated by *Oscillatoria* sp. during seasons.

the increase of Cl content in *H. foetidus* throughout these seasons is caused by its high metabolic activity, as it produces chlorinated compounds during photosynthesis.

#### Chromium in *Hydrurus foetidus*

Great increase of Cr during the autumn will be most probably caused by increased metabolic activity of the algae. Prado *et al.* (2010) described that highest concentrations of Cr during the summer observed

in aquatic fern were caused by increased activity of the plant and its active uptake of the elements. Miranda *et al.* (2012) observed increased accumulation of Cr by living algae than dead. This also suggests that chromium was actively accumulated during the growth of algae.

Sources of Cr can be partially attributed to granodiorite, which contains a small amount of it (Izbicki *et al.* 2008). Other sources of Cr in the area include burning of coal in south Poland which emits chromium particles into the atmosphere (Cheng *et al.* 2014; Molik *et al.* 2018.) Seasonal changes observed in *H. foetidus* are primarily a result of the metabolic needs of the algae, though this is likely not the singular factor.

#### Sulphur and potassium in *Oscillatoria* sp.

Both sulfur and potassium are elements indicating higher biological activity in algae (Subhashini and Kaushik 1986; Ometto 2018; Rashel and Patiño 2019; Ptak *et al.* 2021). Sulfur is one of the main components in algal cells (Giordano 2013) and one of the most important macronutrients for photosynthetic organisms (Giordano and Prioretti 2016). Its main source for algae is  $\text{SO}_4^{2-}$ , which is the most common form of inorganic S in the environment (Giordano and Raven 2014).

Algae show greater dependence on S assimilation

lation from photosynthesis compared to vascular plants (Schmidt 1979). Most S assimilation processes take place in chloroplasts (Brunold and Schiff 1976) where it is crucial for assimilation of cysteine, methionine, and production of dimethylsulphoniopropionate (DMSP) (Giordano and Raven 2014). A study by Rashel and Patiño (2019) found that the algae *Prymnesium parvum* increased its growth rate when exposed to  $\text{MgSO}_4$ . Though  $\text{Mg}^{2+}$  enhanced growth performance,  $\text{SO}_4^{2-}$  stimulated growth of golden alga in a concentration-dependent manner and independently of salinity.

Likens *et al.* (1967) described increased uptake of K during the vegetation period of algae, while Subhashini and Kaushik (1986) found that with increasing concentration of K, protein synthesis and overall biomass of cyanobacteria rose to 0.13 mg K/mL. Photosynthesis is also a potassium dependent process. When algae is deprived of potassium, the volume of produced  $\text{O}_2$  drops drastically, and even with the subsequent addition of potassium, only slight recovery was observed (Clendenning *et al.* 1956; Jackson and Volk 1968).

Exchangeable K varies from < 100 ppm to several thousand ppm. This form of soluble K, available to many organisms, usually constitutes a few ppm of the dry weight of the soil that leaches into the water (Pratt 1965). Its natural sources in this area are mostly derived from the crystalline core of the Tatra Mountains. Primary rocks with high potassium concentrations include quartz, feldspar, and mica (Želazny *et al.* 2011).

Maximum mean concentrations of both elements were measured during the peak biological activity of *Oscillatoria* sp., during summer. Based on these observations, it can be assumed that sulfur and potassium can be a suitable bioindicator of high biological activity and photosynthesis in these species. Seasonal changes in concentrations of these elements are primarily caused by their metabolic needs and not through anthropogenic impact or geochemical processes.

#### Calcium and strontium in *Oscillatoria* sp.

In calcium and strontium, we observed similar behavior. Both decrease during summer and increase throughout autumn and winter. Calcium also experiences large variance in the spring. Snowmelt has been shown to alter concentrations of stream elements in a significant way.  $\text{Ca}^{2+}$  can be stored in snow from mineral dust and is consequently released due to high mobility (Bohleber *et al.* 2018; Avak *et al.* 2019). Other studies have confirmed calcium increases in streams during the spring snowmelt (Stottlemeyer and Troendle 1992; Campbell *et al.* 1995; Moon *et al.* 2014).

However, in recent years, due to increased physical erosion caused by acid rain, weathering of apatite was observed as a rising source of  $\text{Ca}^{2+}$  in the Tatra Mountains (Kopáček *et al.* 2019), and could be washed downstream during snowmelt (Yu *et al.* 2019). Sr and Ca tend to bioaccumulate in strong correlation (Dabbagh *et al.* 2007; Segovia-Campos *et al.* 2023). This correlation was also observed in our research. Strontium does not follow the same pattern during spring, which is indicative

of another, more abundant calcium source during this season. This favors the hypothesis of increased apatite dissolution.

Decreases in Sr and Ca concentrations during summer and increases in colder months can be explained by algal symbionts and their response to sea surface temperatures (SSTs). Sr/Ca ratios in reef coral skeletons are used as a paleothermometer to estimate SSTs. Cohen *et al.* (2002) found that algal symbionts influence Sr/Ca content. Corals with algal symbionts also had inverse correlation with the SSTs but with greater variance during the summer, when the concentration of Sr and Ca are influenced by photosynthesis. Wei *et al.* (2004) discussed that though algal symbionts are more sensitive to SSTs compared to coral, Sr/Ca ratios can yield more precise information about SSTs. It can be suggested that blue-green algae of the order *Oscillatoriales* follows the same pattern. Based on the correlation coefficients ( $r = -0.15 - -0.18$  (Sr);  $-0.24 - -0.37$  (Ca)) developed for US and LS we predict that accumulation of these elements is inversely influenced by the water temperature.

The metabolism of *Oscillatoria* sp. also plays a role also in the accumulation of Sr and Ca as they are vital elements for cell growth and the growth of cell structures (Ray and Bagchi 2002; Dabbagh *et al.* 2007; Shi *et al.* 2013; Sharma 2019; Chen *et al.* 2021; Segovia-Campos *et al.* 2023).

Based on these observations, it can be concluded that the majority of calcium accumulated during spring is released from weathered apatite and mineral dust trapped in snow. Decreases in mean concentration in summer and further increases in autumn and winter is likely driven by sensitivity of the Sr/Ca ratio to changes in water temperature with inverse correlation.

#### Manganese and lead in *Oscillatoria* sp.

Similar results to calcium were observed in manganese and lead. They had the greatest variance (Mn – 115.7 ppm, Pb – 7.2 ppm) during the spring. Changes in discharge and influx from melting snow varied concentrations of these elements. Manganese and lead occur also naturally in the mountains. Manganese is present in the Tatra Mountains in the form of rhodochrosite, whereas manganese oxides (manganite, pyrolusite) (Pawlikowski and Wróbel 2013). During physical weathering it can form  $\text{MnO}_2$  (Ostwald 1988). From these compounds, it can enter streams in the form of manganese cations, which are available to algae (Haug *et al.* 1961; Puranik *et al.* 1999; Das *et al.* 2015). Lead is a trace element naturally occurring in granodiorites, limestones and their soils (Erel *et al.* 1991; Alloway 2012). Its concentrations are regulated by physical weathering (Erel *et al.* 1991; Romanescu *et al.* 2016).

Though snow movement, including avalanches and snowmelt promote bedrock and soil weathering (August *et al.* 2002; Fortner *et al.* 2009) and consequently the release of  $\text{Mn}^{2+}$  and  $\text{Pb}^{2+}$ , we believe, based on previous studies, that the source of these is not only a result of natural sources. Both Mn and Pb are produced during burning of fossil fuels, which can travel long distances (Joselow *et al.* 1978; Janiga 2001; Parmalee and Aschner 2016).

Kopáček *et al.* (2006) and Kwapuliński *et al.* (2012) proved that the source of lead found in Tatra mountain soils was long-range vehicle, industrial, and residential fume emissions. Manganese is also produced by burning gasoline (Joselow *et al.* 1978; Miah *et al.* 2020) and was found by Joselow *et al.* (1978) to be in strong correlation with Pb concentrations.

Large variance of these elements in the spring is likely due to high accumulation of Mn and Pb pollution during late autumn and winter. In the process of snow melting, they are released from the snow and the top layers of soil into the stream as bivalent cations.

#### Chromium in *Oscillatoria* sp.

We observed an increase of Cr in *Oscillatoria* sp. between spring and autumn, where it had exhibited the greatest mean and overall concentrations. During winter, its mean concentration decreased. Different seasonality of Cr uptake by the two species was caused by their differing peaks of biological activity, which greatly contributed to uptake (Prado *et al.* 2010). *Oscillatoria* species were observed by Miranda *et al.* (2012) to accumulate Cr in both its most stable oxidative states, Cr<sup>3+</sup> and Cr<sup>6+</sup>.

Increase of Cr during autumn, when *Oscillatoria* do not experience high metabolic activity is likely due to external factors. This could be caused by emissions from coal burning, which are atmospherically transported and deposited in the Tatra Mountains (Cheng *et al.* 2014; Molik *et al.* 2018). This pollution can be later washed away into the stream.

## References

- Alloway, B.J. (ed.) 2012: Heavy metals in soils. Trace metals and metalloids in soils and their bioavailability. Springer, Dordrecht.
- August, E.E., McKnight, D.M., Hrcir, D.C. and Garhart, K.S. 2002: Seasonal variability of metals transport through a wetland impacted by mine drainage in the Rocky Mountains. *ES&T*, **36**: 3779–3786. <https://doi.org/10.1021/es015629w>
- Avak, S.E., Trachsel, J.C., Edebeli, J., Brüttsch, S., Bartels-Rausch, T., Schneebeli, M., Schwikowski, M. and Eichler A. 2019: Melt-induced fractionation of major ions and trace elements in an Alpine snowpack. *J. Geophys.*, **124**: 1647–1657. <https://doi.org/10.1029/2019JF005026>
- Bohleber, P., Erhardt, T., Spaulding, N., Hoffmann, H., Fischer, H. and Mayewski, P. 2018: Temperature and mineral dust variability recorded in two low-accumulation Alpine ice cores over the last millennium. *Clim. Past*, **14**: 21–37. <https://doi.org/10.5194/cp-14-21-2018>
- Brunold, C. and Schiff, J.A. 1976: Studies of sulfate utilization of algae: 15. Enzymes of assimilatory sulfate reduction in euglena and their cellular localization. *Plant Physiol*, **57**: 430–436. <https://doi.org/10.1104/pp.57.3.430>
- Campbell, D.H., Clow, D.W., Ingersoll, G.P., Mast, M.A., Spahr, N.E. and Turk, J.T. 1995: Processes controlling the chemistry of two snowmelt-dominated streams in the Rocky Mountains. *Water Resour. Res.*, **31**: 2811–2821. <https://doi.org/10.1029/95wr02037>
- Chen, Z., Li, W.Z., Chen, J.Y., Yuan, Z.W. and Chen, X.W. 2021: Effects of calcium ion on the colony formation, growth, and photosynthesis in the edible cyanobacterium *Nostoc sphaeroides*. *J. Appl. Phycol.*, **33**: 1409–1417. <https://doi.org/10.1007/s10811-021-02401-7>
- Cheng, H., Zhou, T., Li, Q., Lu, L. and Lin, C. 2014: Anthropogenic chromium emissions in China from 1990 to 2009. *PLoS one*, **9**: e87753. <https://doi.org/10.1371/journal.pone.0087753>
- Chojnacka, K., Chojnacki, A. and Gorecka, H. 2005: Biosorption of Cr<sup>3+</sup>, Cd<sup>2+</sup> and Cu<sup>2+</sup> ions by blue-green algae *Spirulina* sp.: kinetics, equilibrium and the mechanism of the process. *Chemosphere*, **59**: 75–84. <https://doi.org/10.1016/j.chemosphere.2004.10.005>
- Clendenning, K.A., Brown T.E. and Eyster, H.C. 1956: Comparative studies on photosynthesis in *Nostoc muscorum* and *Chlorella pyrenoidosa*. *Can. J. Bot.*, **34**: 943–966.
- Cohen, A.L., Owens, K.E., Layne, G.D. and Shimizu, N. 2002: The effect of algal symbionts on the accuracy of Sr/Ca paleotemperatures from coral. *Science*, **296**: 331–333. <https://doi.org/10.1126/science.11139-b56-074>
- Cohn, F. 1853: Über lebende Organismen im Trinkwasser. *Zeitschrift kleine Medizin*, **4**: 229–237.
- Dabbagh, R., Ghafourian, H., Baghvand, A., Nabi, G.R., Riahi, H. and Ahmadi Faghih, M.A. 2007: Bioaccumulation and biosorption of stable strontium and 90Sr by *Oscillatoria homogenea* cyanobacterium. *J. Radioanal. Nucl. Ch.*, **272**: 53–59. <https://doi.org/10.1007/s10967-006-6785-4>
- Das, A.P., Ghosh, S., Mohanty, S. and Sukla, L.B. 2015: Advances in manganese pollution and its bioremediation. *Environ. Microbial Biotechnol.*, **45**: 313–328. [https://doi.org/10.1007/978-3-319-19018-1\\_16](https://doi.org/10.1007/978-3-319-19018-1_16)
- Erel, Y., Morgan, J.J. and Patterson, C.C. 1991: Natural levels of lead and cadmium in a remote mountain stream. *Geochim. Cosmochim. Acta*, **55**: 707–719. [https://doi.org/10.1016/0016-7037\(91\)90335-3](https://doi.org/10.1016/0016-7037(91)90335-3)
- Fortner, S.K., Lyons, W.B., Fountain, A.G., Welch, K.A. and Kehrwald, N.M. 2009: Trace element and major ion concentrations and dynamics in glacier snow and melt: Eliot Glacier, Oregon Cascades. *Hydrol. Process.*, **23**: 2987–2996. <https://doi.org/10.1002/hyp.7418>
- Giordano, M. 2013: Homeostasis: an underestimated focal point of ecology and evolution. *Plant Sci.*, **211**: 92–101. <https://doi.org/10.1016/j.plantsci.2013.07.008>
- Giordano, M. and Prioretti, L. 2016: Sulphur and algae: Metabolism, ecology and evolution. In: *The physiology of microalgae* (eds. M. Borowitzka, J. Beardall and J. Raven), pp. 185–209. Developments in Applied Phycology, vol 6, Springer, Cham. [https://doi.org/10.1007/978-3-319-24945-2\\_9](https://doi.org/10.1007/978-3-319-24945-2_9)
- Giordano, M. and Raven, J. A. 2014: Nitrogen and sulfur assimilation in plants and algae. *Aquat. Bot.*, **118**: 45–61. <https://doi.org/10.1016/j.aquabot.2014.06.012>
- Gribble, G.W. 1994: The natural production of chlorinated compounds. *ES&T*, **28**: 310A–319A. <https://doi.org/10.1021/es00056a001>
- Haug, A., Bjerrum, J., Buchardt, O., Olsen, G.E., Pedersen, C. and Toft, J. 1961: The affinity of some divalent metals for different types of alginates. *Acta Chem. Scand.*, **15**: 1794–1795. <https://doi.org/10.3891/acta.chem.scand.15-1794>
- Haug, A. and Smidsrød, O. 1967: Strontium–calcium selectivity of alginates. *Nature*, **215**: 757–757. <https://doi.org/10.1038/215757a0>
- Izbicki, J.A., Ball, J.W., Bullen, T.D. and Sutley, S.J. 2008: Chromium, chromium isotopes and selected trace elements, western Mojave Desert, USA. *Appl. Geochem.*, **23**: 1325–1352. <https://doi.org/10.1016/j.apgeochem.2007.11.015>
- Jackson, W.A. and Volk, R.J. 1968: Role of potassium in photosynthesis and respiration. In: *The role of potassium in agriculture* (eds. V.J. Kilmer, S.E. Younts and N.C. Brady), pp. 109–145. ASA, CSSA, and SSSA. <https://doi.org/10.2134/1968.roleofpotassium.c6>
- Janiga, M. 2001: Birds as bio-indicators of long-transported lead in the Alpine environment. In: *Global change and protected areas* (eds. G. Visconti, M. Beniston, E.D. Iannorelli and D. Barba), pp. 253–259. Advances in Global Change Research, vol 9. Springer, Dordrecht. [https://doi.org/10.1007/0-306-48051-4\\_24](https://doi.org/10.1007/0-306-48051-4_24)



- Joselow, M.M., Tobias, E.D., Koehler, R., Coleman, S., Bogden, J. and Gause, D. 1978: Manganese pollution in the city environment and its relationship to traffic density. *Am. J. Public Health*, **68**: 557–560.
- Kann, E. 1978: Systematik und Ökologie der Algen österreichischer Bergbäche. Vereinigung für theoretische und angewandte Limnologie.
- Klaveness, D. 2019: *Hydnurus foetidus* (Chrysophyceae): an update and request for observations. *Algae*, **34**: 1–5. <https://doi.org/10.4490/algae.2019.34.1.15>
- Kopáček, J., Borovec, J., Hejzlar, J., Kotorová, I., Stuchlík, E. and Veselý, J. 2006: Chemical composition of modern and pre-acidification sediments in the Tatra Mountain lakes. *Biologia*, **61**: 65–76. <https://doi.org/10.2478/s11756-006-0120-y>
- Kopáček, J., Kaňa, J., Bičárová, S., Brahney, J., Navrátil, T., Norton, S.A., Porcal, P. and Stuchlík, E. 2019: Climate change accelerates recovery of the Tatra Mountain lakes from acidification and increases their nutrient and chlorophyll a concentration. *Aquat. Sci.*, **81**: 1–13. <https://doi.org/10.1007/s00027-019-0667-7>
- Küpper, F.C., Carpenter, L.J., McFiggans, G.B., Palmer, C.J., Waite, T.J., Boneberg, E.M., Woitsch, S., Weiler, M., Abela, R., Grolimund, D., Potin, P., Butler, A., Luther, G.W.III, Kroneck, P.M.H. Meyer-Klaucke, W. and Feiters, M.C. 2008: Iodide accumulation provides kelp with an inorganic antioxidant impacting atmospheric chemistry. *PNAS*, **105**: 6954–6958. <https://doi.org/10.1073/pnas.0709959105>
- Kuyucak, N. and Volesky, B. 1989: The mechanism of cobalt biosorption. *Biotechnol. Bioeng.*, **33**: 823–831. <https://doi.org/10.1002/bit.260330705>
- Kwapuliński, J., Pauksto, A., Paprotny, Ł., Musielińska, R., Kowol, J., Nogaj, E. and Rochel, R. 2012: Bioavailability of lead, cadmium, and nickel in Tatra Mountain National Park soil. *Pol. J. Environ. Stud.*, **21**: 407–413.
- La Barre, S., Potin, P., Leblanc, C. and Delage, L. 2010: The halogenated metabolism of brown algae (Phaeophyta), its biological importance and its environmental significance. *Marine Drugs*, **8**: 988–1010. <https://doi.org/10.3390/md8040988>
- Likens, G.E., Bormann, F.H., Johnson, N.M. and Pierce, R.S. 1967: The calcium, magnesium, potassium, and sodium budgets for a small forested ecosystem. *Ecology*, **48**: 772–785. <https://doi.org/10.2307/1933735>
- Lin, Z., Li, J., Luan, Y. and Dai, W. 2020: Application of algae for heavy metal adsorption: A 20-year meta-analysis. *Ecotox. Environ. Safe.*, **190**: 110089. <https://doi.org/10.1016/j.ecoenv.2019.110089>
- Mehta, S.K. and Gaur, J.P. 2001: Characterization and optimization of Ni and Cu sorption from aqueous solution by *Chlorella vulgaris*. *Ecol. Eng.*, **18**: 1–13. [https://doi.org/10.1016/S0925-8574\(00\)00174-9](https://doi.org/10.1016/S0925-8574(00)00174-9)
- Miah, M.R., Ijomone, O.M., Okoh, C.O.A., Ijomone, O.K., Akingbade, G.T., Ke, T., Krum, B., da Cunha Martins Jr., A., Akinyemi, A., Aranoff, N., Soares, F.A.A., Bowman, A.B. and Aschner, M. 2020: The effects of manganese overexposure on brain health. *Neurochem. Int.*, **135**: 104688. [doi:10.1016/j.neuint.2020.104688](https://doi.org/10.1016/j.neuint.2020.104688)
- Miranda, J., Krishnakumar, G. and Gonsalves, R. 2012: Cr<sup>6+</sup> bioremediation efficiency of *Oscillatoria laetevirens* (Crouan & Crouan) Gomont and *Oscillatoria trichoides* Szafer: kinetics and equilibrium study. *J. Appl. Phycol.*, **24**: 1439–1454. <https://doi.org/10.1007/s10811-012-9800-x>
- Molik, A., Trojanowska, M., Łożyńska, M. and Świetlik, R. 2018: Seasonal variations in chromium concentration in urban atmospheric aerosol in the city of Radom. *E3S Web of Conferences*, **28**: 01024. <https://doi.org/10.1051/e3sconf/20182801024>
- Moon, S., Chamberlain, C.P. and Hille, G.E. 2014: New estimates of silicate weathering rates and their uncertainties in global rivers. *Geochim. Cosmochim. Acta*, **134**: 257–274. <https://doi.org/10.1016/j.gca.2014.02.033>
- Nzengung, V.A., Wolfe, L.N., Rennels, D.E., McCutcheon, S.C. and Wang, C. 1999: Use of aquatic plants and algae for decontamination of waters polluted with chlorinated alkanes. *Int. J. Phytoremediation*, **1**: 203–226. <https://doi.org/10.1080/15226519908500016>
- Ometto, F., Steinhovden, K.B., Kuci, H., Lunnbäck, J., Berg, A., Karlsson, A., Handa, A., Wollan, H. and Ejlertsson, J. 2018: Seasonal variation of elements composition and biomethane in brown macroalgae. *Biomass and Bioenergy*, **109**: 31–38. <https://doi.org/10.1016/j.biombioe.2017.11.006>
- Ostwald, J. 1988: Mineralogy of the Groote Eylandt manganese oxides: a review. *Ore Geol. Rev.*, **4**: 3–45. [https://doi.org/10.1016/0169-1368\(88\)90003-0](https://doi.org/10.1016/0169-1368(88)90003-0)
- Palmer, C.M. 1969: A composite rating of algae tolerating organic pollution. *J. Phycol.*, **5**: 78–82. <https://doi.org/10.1111/j.1529-8817.1969.tb02581.x>
- Parmalee, N.L. and Aschner, M. 2016: Manganese and aging. *NeuroToxicology*, **56**: 262–268. <https://doi.org/10.1016/j.neuro.2016.06.006>
- Pawlikowski, M. and Wróbel, M. 2013: Rediscovering old mining activities in the Tatra Mountains. *Geotourism*, **1**: 37–46. <https://doi.org/10.7494/geotour.2013.32-33.37>
- Prado, C., Rosa, M., Pagano, E., Hilal, M. and Prado, F.E. 2010: Seasonal variability of physiological and biochemical aspects of chromium accumulation in outdoor-grown *Salvinia minima*. *Chemosphere*, **81**: 584–593. <https://doi.org/10.1016/j.chemosphere.2010.08.033>
- Pratt, P.F. 1965: Potassium. *Methods of soil analysis: Part 2 chemical and microbiological properties*, **9**: 1022–1030. <https://doi.org/10.2134/agronmonogr9.2.c20>
- Ptak, S.H., Hjulær, A.L., Ditlevsen, S.I., Fretté, X., Errico, M. and Christensen, K.V. 2021: The effect of seasonality and geographic location on sulphated polysaccharides from brown algae. *Aquac. Res.*, **52**: 6235–6243. <https://doi.org/10.1111/are.15485>
- Puranik, P.R., Modak, J.M. and Paknikar, K.M. 1999: A comparative study of the mass transfer kinetics of metal biosorption by microbial biomass. *Hydrometallurgy*, **52**: 189–198. [https://doi.org/10.1016/S0304-386X\(99\)00017-1](https://doi.org/10.1016/S0304-386X(99)00017-1)
- Rashel, R.H. and Patiño, R. 2019: Growth response of the ichthyotoxic haptophyte, *Prymnesium parvum* Carter, to changes in sulfate and fluoride concentrations. *PLoS one*, **14**: e0223266. <https://doi.org/10.1371/journal.pone.0223266>
- Ray, S. and Bagchi, S.N. 2002: Nutrients and pH regulate algicide accumulation in cultures of the cyanobacterium *Oscillatoria laetevirens*. *New Phytol.*, **149**: 455–460. <https://doi.org/10.1046/j.1469-8137.2001.00061.x>
- Romanescu, G., Miftode, D., Pintilie, A.M., Stoleriu, C.C. and Sandu, I. 2016: Water quality analysis in mountain freshwater: Poiana Uzului Reservoir in the Eastern Carpathians. *Rev. Chim.(Bucharest)*, **67**: 2318–2326.
- Roy, A., Gogoi, N., Yasmin, F. and Farooq, M. 2022: The use of algae for environmental sustainability: Trends and future prospects. *Environ. Sci. Pollut. Res.*, **29**: 40373–40383. <https://doi.org/10.1007/s11356-022-19636-7>
- Salama, E.S., Roh, H.S., Dev, S., Khan, M.A., Abou-Shanab, R.A.I., Chang, S.W. and Jeon, B.H. 2019: Algae as a green technology for heavy metals removal from various wastewater. *World J. Microbiol. Biotechnol.*, **35**: 1–19. <https://doi.org/10.1007/s11274-019-2648-3>
- Schmidt, A. 1979: Photosynthetic assimilation of sulphur compounds. In: *Photosynthesis II Encyclopedia of plant physiology* (eds. M.L. Gibbs and E. Latzko), pp. 481–496. Springer, Berlin, Heidelberg. [https://doi.org/10.1007/978-3-642-67242-2\\_37](https://doi.org/10.1007/978-3-642-67242-2_37)
- Segovia-Campos, I., Filella, M., Perron, K. and Ariztegui, D. 2023: High calcium and strontium uptake by the green microalga *Tetraselmis chui* is related to micropeal formation and cell growth. *Environ. Microbiol. Rep.*, **15**: 38–50. <https://doi.org/10.1111/1758-2229.13124>
- Seki, H. and Suzuki, A. 1998: Biosorption of heavy metal ions to brown algae, *Macrocystis pyrifera*, *Kjellmaniella crassifolia*, and *Undaria pinnatifida*. *J. Colloid*

- Interface Sci.*, **206**: 297–301. <https://doi.org/10.1006/jcis.1998.5731>
- Sharma, S. 2019: Uptake, transport, and remediation of strontium. In: *Strontium contamination in the environment*. (eds. P. Pathak and D. Gupta), pp. 99–119. The Handbook of Environmental Chemistry, vol 88. Springer, Cham. [https://doi.org/10.1007/978-3-030-15314-4\\_6](https://doi.org/10.1007/978-3-030-15314-4_6)
- Shi J.Q., Wu Z.X. and Song L.R. 2013: Physiological and molecular responses to calcium supplementation in *Microcystis aeruginosa* (Cyanobacteria). *N. Z. J. Mar. Freshwater Res.*, **47**: 51–61. <https://doi.org/10.1080/00288330.2012.741067>
- Stottleyer, R. and Troendle, C.A. 1992: Nutrient concentration patterns in streams draining alpine and subalpine catchments, Fraser Experimental Forest, Colorado. *J. Hydrol.*, **140**: 179–208. [https://doi.org/10.1016/0022-1694\(92\)90240-V](https://doi.org/10.1016/0022-1694(92)90240-V)
- Subhashini, D. and Kaushik, B.D. 1986: Uptake of sodium and potassium by blue-green algae (Cyanobacteria). *Zentralblatt für Mikrobiologie*, **141**: 177–180. [https://doi.org/10.1016/S0232-4393\(86\)80055-5](https://doi.org/10.1016/S0232-4393(86)80055-5)
- Wehr, J.D. and Sheath, R.G. 2003: Freshwater habitats of algae. In: *Freshwater Algae of North America – Ecology and Classification* (eds. J.D. Wehr and R.G. Sheath), pp. 11–57. Academic Press, San Diego, Burlington. <https://doi.org/10.1016/B978-0-12-385876-4.00002-5>
- Wei, G., Yu, K. and Zhao, J. 2004: Sea surface temperature variations recorded on coralline Sr/Ca ratios during Mid-Late Holocene in Leizhou Peninsula. *Chin. Sci. Bull.*, **49**: 1876–1881. <https://doi.org/10.1007/bf03183416>
- Yu, Z., Wu, G., Keys, L., Li, F., Yan, N., Qu, D. and Liu, X. 2019: Seasonal variation of chemical weathering and its controlling factors in two alpine catchments, Nam Co basin, central Tibetan Plateau. *J. Hydrol.*, **576**: 381–395. <https://doi.org/10.1016/j.jhydrol.2019.06.042>
- Želazny, M., Astel, A., Wolanin, A. and Małek, S. 2011: Spatiotemporal dynamics of spring and stream water chemistry in a high-mountain area. *Environ. Pollut.*, **159**: 1048–1057. <https://doi.org/10.1016/j.envpol.2010.11.021>
- Zeraatkar, A.K., Ahmadzadeh, H., Talebi, A.F., Mohemani, N.R. and McHenry, M.P. 2016: Potential use of algae for heavy metal bioremediation, a critical review. *J. Environ. Manage.*, **181**: 817–831. <https://doi.org/10.1016/j.jenvman.2016.06.059>

Received 23 April 2024; accepted 5 June 2024.